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1 Sorption, microbial incorporation, and decomposition of position-specific ¹⁴C-labeled acetate in soil

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Low molecular organic acids (LMOA) are released into soil by root exudation or plant and microorganisms residue decomposition. They are subject to a variety of processes: microbial utilization and decomposition to CO_2 , sorption by mineral particles and soil organic matter, or leaching. Among LMOA acetate is most abundant and occurs typically in concentrations from 0.5 to 1000 μ M in soil solution.

All previous studies to transformation of LMOA in soil used uniformly ¹⁴C or ¹³C labeled substances. The amount of ¹⁴C or ¹³C served as parameter of substance concentration in solution, its sorption, and decomposition of the whole molecule. With these approaches it remained unclear which functional group (in the case of acetate CH₃- or COOH-) controlled the processes and whether it was the whole molecule or one of its metabolites. To overcome this shortcoming and to trace C from individual positions in the molecule we conducted experiment with 1-¹⁴C-acetate and 2-¹⁴C-acetate. By this position specific labeling we distinguished between C from the CH₃-group and C from the COOH-group of the molecule contributing to all processes. When results for these two ¹⁴C positions differ from each other it is probable that the two functional

groups are subject to different fates in the respective process.

Acetate was added to 1 g of dried, sieved, and re-moistened loamy soil as 2.5 ml aqueous solution and gently shaken for periods between 10 s and 1 d. Concentrations in the soil solution varied from 0.1 to 1000 μ mol l⁻¹. ¹⁴C-activity in the supernatant was measured after centrifugation at 1500 g and represented not absorbed acetate. Sterilization by CHCl₃ or HgCl₂ + NaN₃ allowed separate determination of microbial decomposition and sorption to the soil matrix.

The equilibrium between sorption, incorporation, decomposition to CO_2 was reached after 28 min for ¹⁴C from CH₃- and after 85 min for ¹⁴C from COOH-. In the not sterilized samples ¹⁴C from CH₃-group declined in soil solution faster than ¹⁴C from COOH-group. Higher ¹⁴C recovery in soil from CH₃- than from COOH- indicated that CH₃- was preferentially incorporated in microorganisms. The losses from the soil-solution system (= decomposition to CO_2) were equal for ¹⁴C from both groups.

In CHCl₃ sterilized soil samples the sorption was influenced by substances released from microbial cells. However, in HgCl₂/NaN₃-sterilized samples sorption of ¹⁴C from both groups did not differ significantly.

After combining the information from sterile and not sterile conditions, the following pool sizes could be determined for acetate at 100 μ M: of the COOH-group 15.9 \pm 0.9% were sorbed to the soil matrix, 47.8 \pm 0.9% were decomposed to CO₂, and 22.8 \pm 0.7% remained in solution. For the CH₃ was found: sorption 15.6 \pm 0.5%, decomposition to CO₂ 47.1 \pm 0.3%, solution 13.6 \pm 0.4%. Thus, about 24% of CH₃ was incorporated into microorganisms compared to only about 13% of COOH.

At concentrations higher than 100 μ M and incorporation and decomposition decreased. For CH₃ at 1000 μ M decomposition was 10.8±0.9%, for COOH 20.5±5.5%. Obviously, allotted time (200 min) was too short for complete decomposition.

With the presented approach we showed that the functional groups of acetate are subject to different processes in soil. The use of position-specific labeled substances is an appropriate tool to elucidate these differences.